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# Synthesis of carba-analogues of myoseverin by regioselective cross-coupling reactions of 2,6-dichloro-9-isopropylpurine

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Abstract—A series of 9-isopropylpurine derivatives bearing 4-methoxyphenyl, 4-methoxybenzyl, (4-methoxyphenyl)ethynyl and 2-(4-methoxyphenyl)ethyl groups in positions 2 and 6 were prepared as carba-analogues of antimitotic myoseverin. Cross-coupling reactions of 2,6-dichloro-9-isopropylpurine (1) with one equivalent of (4-methoxyphenyl)boronic acid or (4-methoxybenzyl)zinc chloride gave regioselectively the 6-substituted 2-chloropurines which were used for another cross-coupling reaction with a second equivalent of the organometallic reagent. The Sonogashira reaction of 1 with 4-(methoxyphenyl)ethyne gave 2,6-bis[(4-methoxyphenyl)ethynyl]-9-isopropylpurine that was hydrogenated to 2,6-bis[2-(4-methoxyphenyl)ethyl]-9-isopropylpurine. Regioselectivity of the couplings was proved by means of  ${}^{1}H-{}^{15}N$  HMBC experiments. 2,6-Bis[(4-methoxyphenyl)ethynyl]-9-isopropylpurine showed considerable cytostatic activity, while the other compounds were inactive. © 2003 Elsevier Science Ltd. All rights reserved.

### 1. Introduction

In the last decade, several biological activities of 9-alkyl-2,6-bis(alkyl or benzylamino)purines were reported: inhibition of CDK,<sup>1</sup> estrogen sulfotransferase<sup>2</sup> and inositol-1,4,5-triphosphate-3-kinase.<sup>3</sup> Recently, a novel cytostatic compound myoseverin, 2,6-bis[(4-methoxybenzyl)amino]-9-isopropylpurine, has been discovered by screening of a purine library.<sup>4</sup> This compound blocks tubulin polymerization with moderate activity. Later on, some 9-cycloalkyl derivatives<sup>5</sup> as well as analogously substituted triazines<sup>6</sup> were found to exhibit a somewhat better activity. Myoseverin also affects cultures of hybridoma cells producing monoclonal antibodies.<sup>7</sup>

6-Aryl-,<sup>8</sup> 6-benzyl-<sup>9</sup> and 6-alkynylpurine<sup>10</sup> derivatives were also recently reported to possess cytostatic activity. A combination of the structural features of these two classes of antineoplastic compounds led us to the design of carbaanalogues of myoseverin (Fig. 1) consisting of 9-isopropylpurine derivatives bearing 4-methoxyphenyl, 4-methoxybenzyl, (4-methoxyphenyl)ethynyl and isosteric 2-(4-methoxyphenyl)ethyl groups in the positions 2 and 6. The replacement of the (4-methoxybenzyl)amino groups by C–C linked 4-methoxyphenyl groups should prevent catabolic degradation by deaminases and also test the role of the NH group in the binding with a biological target; on the other hand it will definitely decrease water-solubility of the compounds. The synthesis of these carba-analogues is the subject of this paper.

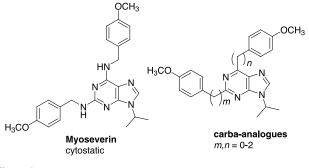


Figure 1.

## 2. Results and discussion

In order to prepare the abbreviated analogues, regioselective cross-coupling reactions<sup>11</sup> of 2,6-dichloropurines have been employed.<sup>12</sup> Thus the reaction of 2,6-dichloro-9-isopropylpurine (1)<sup>13</sup> with one equivalent of (4-methoxyphenyl)boronic acid gave selectively the substitution at position 6 to form 2-chloro-6-(4-methoxyphenyl)purine 2 in 83% yield. Analogously the reaction of 2,6-dichloropurine 1 with one equivalent of (4-methoxybenzyl)zinc chloride gave regioselectively 2-chloro-6-(4-methoxybenzyl)purine 3 in 76%

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yield. Another cross-coupling reaction of **2** with (4-methoxybenzyl)zinc chloride or of **3** with (4-methoxyphenyl)boronic acid furnished the heterodisubstituted purines **4** or **5** in 91 and 96% yields, respectively. Cross-coupling of **1** with three equivalents of (4-methoxyphenyl)boronic acid or (4-methoxybenzyl)zinc chloride afforded the 2,6-bis(4methoxyphenyl or 4-methoxybenzyl)purines **6** and **7** in high yields of 94 and 90%, respectively (Scheme 1).

The Sonogashira cross-coupling reaction of 1 with 2 equiv. of (4-methoxyphenyl)acetylene gave the 2,6-bis[(4-methoxyphenyl)ethynyl]purine 8 in a moderate yield of 39%. Catalytic hydrogenation of 8 on Pd/C afforded the 2,6-bis(4-methoxyphenethyl)purine 9 in a low isolated yield of 14% accompanied by a complex mixture of partly hydrogenated and oligo/polymeric by-products. The yield of 9 could probably be improved by optimization of conditions and selection of other catalysts (Pt or Ni). Nevertheless, despite the lower yields, the compounds 8 and 9 were prepared in sufficient amounts and purity for the biological activity screening.

Though the cross-coupling reactions of 2,6-dichloro-9isopropylpurine (**1**) should follow the same regioselectivity as reported earlier,<sup>12</sup> we have decided to make an independent proof by direct NMR methods. In order to distinguish between the 2-benzyl-6-phenylpurine **4** and 6benzyl-2-phenylpurine **5** derivatives, standard  ${}^{1}\text{H}{-}{}^{13}\text{C}$ HMBC experiments were used. However, the crucial cross-peaks affording three-bond correlations between the CH<sub>2</sub> and C-5 were overlapped with aromatic signals. Therefore, for the assignment of these compounds we utilized  ${}^{1}\text{H}{-}{}^{15}\text{N}$  HMBC experiments (Table 1, Fig. 2). Using this standard procedure optimized for long-range couplings of 7–10 Hz (50–60 ms delay) two- and threebond correlations were observed. The assignment of compound **4** was based on the presence of cross-peaks between methylene protons and nitrogens N-1 and N-3,

Table 1.  $^{15}N$  NMR chemical shifts and  $^{3}(^{1}H,^{15}N)$  connectivities from  $^{1}H-^{15}N$  HMBC spectra of compounds 4, 5 and 7 in CDCl<sub>3</sub> at 298 K

Compound	<sup>15</sup> N shifts, $\delta$				<sup>3</sup> ( <sup>1</sup> H, <sup>15</sup> N) connectivities	
	N-1	N-3	N-7	N-9	2-CH <sub>2</sub> Ar	6-C <b>H</b> ₂Ar
4	267.3	242.9	238.7	172.7	N-1, N-3	_
5	270.6	n.o. <sup>a</sup>	236.8	174.7	_	N-1
7	277.4	245.6	237.7	174.6	N-1, N-3	N-1

<sup>a</sup> Not observed.

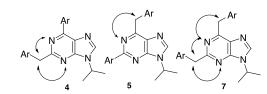
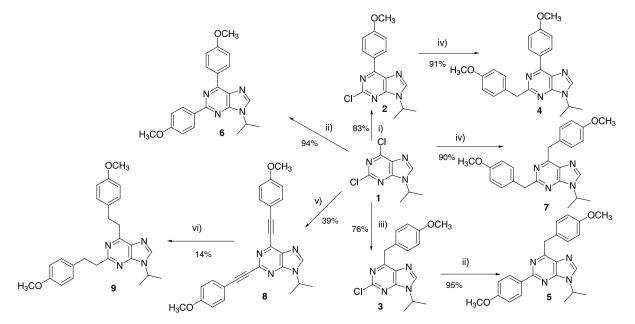


Figure 2. <sup>1</sup>H-<sup>15</sup>N HMBC connectivities in compounds 4, 5 and 7.

while in the case of compound **5** only one cross-peak, indicating a connectivity between methylene protons and nitrogen N-1 was observed. Accordingly, in  ${}^{1}\text{H}{-}{}^{15}\text{N}$  HMBC spectrum of compound **7** we found three-bond correlation signals between both methylene groups and corresponding nitrogens.

In conclusion, abbreviated carba-analogues of myoseverin were regioselectively and efficiently prepared by crosscoupling reactions of 2,6-dichloro-9-isopropylpurine (1) with (4-methoxyphenyl)boronic acid and/or (4-methoxybenzyl)zinc chloride. Much less efficient was the coupling of 1 with (4-methoxyphenyl)acetylene followed by catalytic hydrogenation leading to 2,6-bis(4-methoxyphenethyl)purine 9.

The target compounds 2-9 were tested on their in vitro inhibition of the cell growth in the following cell cultures:



Scheme 1. (i) 4-MeOC<sub>6</sub>H<sub>4</sub>B(OH)<sub>2</sub> (1 equiv.), K<sub>2</sub>CO<sub>3</sub>, Pd(PPh<sub>3</sub>)<sub>4</sub>; (ii) 4-MeOC<sub>6</sub>H<sub>4</sub>B(OH)<sub>2</sub> (2 equiv.), K<sub>2</sub>CO<sub>3</sub>, Pd(PPh<sub>3</sub>)<sub>4</sub>; (iii) 4-MeOC<sub>6</sub>H<sub>4</sub>CH<sub>2</sub>ZnCl (1 equiv.), K<sub>2</sub>CO<sub>3</sub>, Pd(PPh<sub>3</sub>)<sub>4</sub>; (iv) 4-MeOC<sub>6</sub>H<sub>4</sub>CH<sub>2</sub>ZnCl (2 equiv.), K<sub>2</sub>CO<sub>3</sub>, Pd(PPh<sub>3</sub>)<sub>4</sub>; (v) 4-CH<sub>3</sub>OC<sub>6</sub>H<sub>4</sub>C $\equiv$ CH, CuI, Et<sub>3</sub>N, Pd(PPh<sub>3</sub>)<sub>4</sub>, DMF; (vi) H<sub>2</sub>, Pd/C, dioxane/EtOH.

mouse leukemia L1210 cells (ATCC CCL 219); human promyelocytic leukemia HL60 cells (ATCC CCL 240); human cervix carcinoma HeLa S3 cells (ATCC CCL 2.2); and human T lymphoblastoid CCRF-CEM cell line (ATCC CCL 119). The results showed that the 2,6-bis[(4-methoxyphenyl)ethynyl]purine **8** exhibits considerable activity ( $IC_{50}=0.6$ , 1.6 and 3.1  $\mu$ M against L1210, HL60 and CCRF-CEM cell-lines, respectively; for comparison: myoseverin<sup>6</sup>  $IC_{50}=10 \ \mu$ M for U937 cell line). Compound **5** showed a moderate activity against L1210 ( $IC_{50} \sim 5 \ \mu$ M—low water solubility), while the other compounds of this series were inactive ( $IC_{50} > 20 \ \mu$ M) in these assays. The lack of activity in most of the compounds could be caused by the lower water solubility and/or limited transport into the cells.

#### 3. Experimental

Melting points were determined on a Kofler block and are uncorrected. NMR spectra were measured on a Bruker AMX3 400 (<sup>1</sup>H, 400.13 MHz and <sup>13</sup>C, 100.62 MHz) or a Bruker DRX 500 Avance (<sup>1</sup>H, 500.13 MHz and <sup>13</sup>C, 125.77 MHz) spectrometer at 298 K. Standard <sup>1</sup>H-<sup>15</sup>N HMBC experiments were recorded in CDCl<sub>3</sub> using a Bruker Avance DRX 500 spectrometer operating at frequencies of 500.13 MHz (<sup>1</sup>H) and 50.68 MHz ( $^{15}$ N).  $^{15}$ N NMR resonances were referenced to the signal of liquid CH<sub>3</sub>NO<sub>2</sub> (381.7 ppm). Unambiguous assignment of the NMR signals is based on <sup>13</sup>C{<sup>1</sup>H}, <sup>13</sup>C APT, COSY and  $^{1}H-^{13}C$  and  $^{1}H-^{15}N$  HMBC spectra. IR spectra were recorded on Nicolet 750 FT-IR. Mass spectra were measured on ZAB-SEQ (VG Analytical). Microanalyses were performed on a Perkin-Elmer 240-II CHN Analyser. Silica gel (ICN SiliTech, 32-63) was used for column chromatography. Toluene was degassed in vacuo and stored over molecular sieves under argon. DMF was distilled from P<sub>2</sub>O<sub>5</sub>, degassed in vacuo and stored over molecular sieves under argon. THF was refluxed with Na and benzophenone under argon and freshly distilled prior to use. (4-Methoxyphenyl)boronic acid and Rieke® (4-methoxybenzyl)zinc chloride were supplied by Aldrich. Starting compound 1 was prepared according to known<sup>13</sup> procedure. Cytostatic activity tests were performed as described in Ref. 8.

## 3.1. Cross-coupling reactions with (4-methoxyphenyl)boronic acid—general procedure A

Toluene (10 ml) was added to an argon-purged flask containing the chloropurine (1 mmol),  $K_2CO_3$  (200 mg, 1.5 mmol), (4-methoxyphenyl)boronic acid (1 or 3 mmol) and Pd(PPh<sub>3</sub>)<sub>4</sub> (59 mg, 0.05 mmol) and the mixture was stirred under argon at 100°C for 8 h. After cooling to ambient temperature the solvent was evaporated in vacuo and the residue was chromatographed on a silica gel column (100 g, ethyl acetate–light petroleum 1:2 to 9:1). Evaporation and drying of the product containing fractions afforded the (4-methoxyphenyl)purines as foams or amorphous solids.

## **3.2.** Cross-coupling reactions with (4-methoxybenzyl)zinc chloride—general procedure B

THF (10 ml) was added to an argon-purged flask containing

the chloropurine (1 mmol) and  $Pd(PPh_3)_4$  (59 mg, 0.05 mmol). The mixture was stirred at ambient temperature for 10 min and, after dissolution of the solids, a solution of (4-methoxybenzyl)zinc chloride (Rieke® organozinc reagent, 0.5 M solution in THF, 2 or 6 ml, 1 or 3 mmol) was added dropwise (within 10 min) at ambient temperature. Stirring at ambient temperature was continued for 15 min followed by stirring at 60°C for 8 h. Then the reaction mixture was allowed to stand overnight at ambient temperature and poured into saturated aqueous NH<sub>4</sub>Cl (10 ml). To this mixture, saturated aqueous Na<sub>2</sub>EDTA (10 ml) was added and the mixture was stirred for 10 min. Then the reaction mixture was extracted with ethyl acetate  $(3\times 20 \text{ ml})$  and the collected organic layers were washed with saturated aqueous Na<sub>2</sub>EDTA (20 ml) and brine (20 ml), dried with anhydrous MgSO<sub>4</sub> and evaporated in vacuo. Column chromatography of the residue on silica gel (100 g, ethyl acetate-light petroleum 1:2 to 9:1) afforded, after evaporation and drying, the (4-methoxybenzyl)purines as amorphous solids.

3.2.1. 2-Chloro-9-isopropyl-6-(4-methoxyphenyl)purine (2). Prepared from 1 by procedure A (1 equiv. of boronic acid) in 83%. Colorless crystals, mp 210-214°C (CH<sub>2</sub>Cl<sub>2</sub>/ heptane). EI MS, m/z: 302 (78) [M], 260 (100). IR (KBr),  $\nu = 1607, 1586, 1570, 1518, 1460, 1405, 1325 \text{ cm}^{-1}$ . <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): 1.64 (d, 6H, J=6.7 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 3.90 (s, 3H, OCH<sub>3</sub>); 4.95 (sept., 1H, J=6.7 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 7.05 (d, 2H, J=8.7 Hz, H-arom.); 8.13 (s, 1H, H-8); 8.82 (d, 2H, J=8.7 Hz, H-arom.). <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>): 22.62 (CH(CH<sub>3</sub>)<sub>2</sub>); 47.25 (CH(CH<sub>3</sub>)<sub>2</sub>); 55.41 (OCH<sub>3</sub>); 114.10 and 131.93 (CHarom.); 127.39 (C-i-arom.); 129.81 (C-5); 141.89 (CH-8); 153.55, 153.89 and 156.22 (C-2, C-4 and C-6); 162.56 (C-OMe). Exact mass (EI HR MS) found: 302.0943; for C<sub>15</sub>H<sub>15</sub>ClN<sub>4</sub>O calculated: 302.0934. Anal. calcd for C<sub>15</sub>H<sub>15</sub>ClN<sub>4</sub>O (302.8): C, 59.50; H, 4.99; N, 18.51; found: C, 59.50; H, 4.97; N, 18.40.

**3.2.2. 2-Chloro-9-isopropyl-6-(4-methoxybenzyl)purine** (3). Prepared from 1 by procedure B (1 equiv. of organozinc reagent) in 76%. Colorless oil. EI MS, *m/z*: 316 (18) [M], 274 (10), 259 (12), 121 (8), 28 (100). IR (CHCl<sub>3</sub>),  $\nu$ =1552, 1483, 1435, 1348, 1206 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 1.60 (d, 6H, *J*=6.8 Hz, CH(*CH*<sub>3</sub>)<sub>2</sub>); 3.75 (s, 3H, OCH<sub>3</sub>); 4.41 (s, 2H, CH<sub>2</sub>); 4.87 (sept., 1H, *J*=6.8 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 6.82 (d, 2H, *J*=8.6 Hz, H-arom.); 7.42 (d, 2H, *J*=8.6 Hz, H-arom.); 8.09 (s, 1H, H-8). <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>): 22.50 (CH(*C*H<sub>3</sub>)<sub>2</sub>); 38.68 (CH<sub>2</sub>); 47.46 (CH(CH<sub>3</sub>)<sub>2</sub>); 55.18 (OCH<sub>3</sub>); 113.99 and 130.35 (CH-arom.); 129.28 (C*-i*-arom.); 131.74 (C-5); 142.36 (CH-8); 152.45, 153.75 and 158.50 (C-2, C-4 and C-6); 163.08 (*C*-OMe). Exact mass (EI HR MS) found: 316.1084; for C<sub>16</sub>H<sub>17</sub>ClN<sub>4</sub>O calculated: 316.1091.

**3.2.3. 9-Isopropyl-2-(4-methoxybenzyl)-6-(4-methoxyphenyl)purine (4).** Prepared from **2** by procedure B (1.5 equiv. of organozinc reagent) in 91%. Colorless oil. EI MS, *m/z*: 388 (61) [M], 345 (20), 277 (15), 149 (100). IR (KBr),  $\nu$ =1608, 1577, 1513, 1464, 1391, 1372, 1302, 1253 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): 1.62 (d, 6H, *J*=6.7 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 3.77 and 3.88 (2×s, 2×3H, 2×OCH<sub>3</sub>); 4.33 (s, 2H, CH<sub>2</sub>); 4.94 (sept., 1H, *J*=6.7 Hz,

CH(CH<sub>3</sub>)<sub>2</sub>); 6.84 (d, 2H, J=8.2 Hz, H-arom.); 7.05 (d, 2H, J=8.4 Hz, H-arom.); 7.42 (d, 2H, J=8.2 Hz, H-arom.); 8.07 (s, 1H, H-8); 8.78 (d, 2H, J=8.4 Hz, H-arom.). <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>): 22.57 (CH(CH<sub>3</sub>)<sub>2</sub>); 45.15 (CH<sub>2</sub>); 46.79 (CH(CH<sub>3</sub>)<sub>2</sub>); 55.21 and 55.33 (2×OCH<sub>3</sub>); 113.67, 113.91, 130.15 and 131.43 (CH-arom.); 128.93, 128.99 and 131.68 (2×C-*i*-arom. and C-5); 140.96 (CH-8); 152.64, 154.06 and 158.09 (C-2, C-4 and C-6); 161.72 and 163.39 (2×C-OMe). Exact mass (EI HR MS) found: 388.1907; for C<sub>23</sub>H<sub>24</sub>N<sub>4</sub>O<sub>2</sub> calculated: 388.1899.

3.2.4. 9-Isopropyl-6-(4-methoxybenzyl)-2-(4-methoxyphenyl)purine (5). Prepared from 3 by procedure A (1.5 equiv. of boronic acid) in 95%. Colorless oil. EI MS, m/z: 388 (100) [M], 345 (45), 277 (15), 149 (40). IR (KBr),  $\nu$ =1610, 1593, 1511, 1494, 1465, 1438, 1389, 1375, 1302, 1250 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): 1.66 (d, 6H, J=6.8 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 3.75 and 3.88 (2×s, 2×3H, 2×OCH<sub>3</sub>); 4.47 (s, 2H, CH<sub>2</sub>); 4.95 (sept., 1H, J=6.8 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 6.83 (d, 2H, J=8.6 Hz, H-arom.); 7.00 (d, 2H, J=8.8 Hz, H-arom.); 7.48 (d, 2H, J=8.6 Hz, H-arom.); 8.03 (s, 1H, H-8); 8.50 (d, 2H, J=8.8 Hz, H-arom.). <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>): 22.56 (CH(CH<sub>3</sub>)<sub>2</sub>); 38.76 (CH<sub>2</sub>); 47.12 (CH(CH<sub>3</sub>)<sub>2</sub>); 55.17 and 55.34 (2×OCH<sub>3</sub>); 113.68, 113.79, 129.79 and 130.36 (CH-arom.); 130.41, 130.86 and 131.26 (2×C-i-arom. and C-5); 141.46 (CH-8); 151.69 (C-4); 158.19, 158.44, 160.29 and 161.19 (C-2, C-6, 2×C-OMe). Exact mass (EI HR MS) found: 388.1911; for C<sub>23</sub>H<sub>24</sub>N<sub>4</sub>O<sub>2</sub> calculated: 388.1899.

3.2.5. 9-Isopropyl-2,6-bis(4-methoxyphenyl)purine (6). Prepared from 1 by procedure A (3 equiv. of boronic acid) in 94%. Colorless crystals, mp 175-176°C (CH<sub>2</sub>Cl<sub>2</sub>/heptane). EI MS, m/z: 374 (80) [M], 332 (59), 149 (100). IR (KBr),  $\nu$ =1609, 1588, 1565, 1513, 1433, 1372, 1246 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): 1.64 (d, 6H, J=6.6 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 3.84 and 3.85 (2×s, 2×3H, 2×OCH<sub>3</sub>); 4.97 (sept., 1H, J=6.6 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 6.98 (d, 2H, J=8.5 Hz, H-arom.); 7.04 (d, 2H, J=8.5 Hz, H-arom.); 8.03 (s, 1H, H-8); 8.56 (d, 2H, J=8.5 Hz, H-arom.); 8.88 (d, 2H, J=8.5 Hz, H-arom.). <sup>13</sup>C NMR (100.6 MHz. CDCl<sub>3</sub>): 22.59  $(CH(CH_3)_2);$ 47.00 (CH(CH<sub>3</sub>)<sub>2</sub>); 55.34 (2×OCH<sub>3</sub>); 113.69, 113.90, 129.77 and 131.43 (CH-arom.); 129.19 and 129.31 (2×C-i-arom.); 131.50 (C-5); 141.28 (CH-8); 152.96, 153.72 and 158.02 (C-2, C-4 and C-6); 161.24 and 161.75 (2×C-OMe). Exact mass (EI HR MS) found: 374.1754; for C<sub>22</sub>H<sub>22</sub>N<sub>4</sub>O<sub>2</sub> calculated: 374.1743. Anal. calcd for C<sub>22</sub>H<sub>22</sub>N<sub>4</sub>O<sub>2</sub> (374.4): C, 70.57; H, 5.92; N, 14.96; found: C, 70.56; H, 5.92; N, 14.95.

**3.2.6. 9-Isopropyl-2,6-bis(4-methoxybenzyl)purine** (7). Prepared from **1** by procedure B (3 equiv. of organozinc reagent) in 90%. Colorless oil. EI MS, *m/z*: 402 (100) [M], 359 (34), 345 (11), 149 (21). IR (KBr),  $\nu$ =1611, 1594, 1513, 1496, 1464, 1442, 1391, 1382, 1325, 1301, 1249 cm<sup>-1</sup>. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): 1.58 (d, 6H, *J*=6.7 Hz, CH(*CH*<sub>3</sub>)<sub>2</sub>); 3.74 and 3.77 (2×s, 2×3H, 2×OCH<sub>3</sub>); 4.27 and 4.40 (2×s, 2×2H, 2×CH<sub>2</sub>); 4.85 (sept., 1H, *J*=6.7 Hz, *CH*(CH<sub>3</sub>)<sub>2</sub>); 6.79 (d, 2H, *J*=9.0 Hz, H-arom.); 6.81 (d, 2H, *J*=8.9 Hz, H-arom.); 7.32 (d, 2H, *J*=8.3 Hz, H-arom.); 7.39 (d, 2H, *J*=8.3 Hz, H-arom.); 8.00 (s, 1H, H-8). <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>): 22.51 (CH(*C*H<sub>3</sub>)<sub>2</sub>); 38.65 and 44.93 (2×CH<sub>2</sub>); 46.95 (*C*H(CH<sub>3</sub>)<sub>2</sub>); 55.17 and 55.22 (2×OCH<sub>3</sub>); 113.63, 113.78, 130.11 and 130.32 (CH-arom.); 130.73,

131.50 (2×C-*i*-arom. and C-5); 141.19 (CH-8); 151.49, 158.11 and 158.23 (C-2, C-4 and C-6); 160.50 and 163.71 (2×C-OMe). Exact mass (EI HR MS) found: 402.2064; for  $C_{24}H_{26}N_4O_2$  calculated: 402.2056. Anal. calcd for  $C_{24}H_{26}N_4O_2$  (402.5): C, 71.62; H, 6.51; N, 13.92; found: C, 71.24; H, 6.58; N, 13.63.

3.2.7. 9-Isopropyl-2,6-bis[(4-methoxyphenyl)ethynyl]purine (8). DMF (5 ml) and Et<sub>3</sub>N (2 ml) were added through septum to an argon purged flask containing 2,6dichloropurine 1 (690 mg, 3 mmol), (4-methoxyphenyl)acetylene<sup>14</sup> (1.32 g, 10 mmol), CuI (200 mg, 1 mmol) and  $Pd(PPh_3)_4$  (200 mg, 0.174 mmol). The mixture was then stirred at 120°C for 12 h and left overnight at ambient temperature. The solvents were evaporated in vacuo and the residue was chromatographed on a silica gel column (150 g, ethyl acetate-light petroleum 1:2) to give compound  $\mathbf{8}$  as yellow amorphous solid (500 mg, 39%). FAB MS, m/z: 423 (20) [M], 279 (18), 135 (100). IR (CHCl<sub>3</sub>), v=2211, 1605, 1569, 1512, 1487, 1465, 1379, 1295, 1252 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 1.66 (d, 6H, *J*=6.8 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 3.84 and 3.85 (2×s, 2×3H, 2×OCH<sub>3</sub>); 5.03 (sept., 1H, J=6.8 Hz,  $CH(CH_3)_2$ ; 6.90 (d, 2H, J=8.4 Hz, H-arom.); 6.92 (d, 2H, J=8.2 Hz, H-arom.); 7.65 (d, 2H, J=8.7 Hz, H-arom.); 7.71 (d, 2H, J=8.7 Hz, H-arom.); 8.24 (s, 1H, H-8). <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>): 22.70 (CH(CH<sub>3</sub>)<sub>2</sub>); 47.21 (CH(CH<sub>3</sub>)<sub>2</sub>); 55.28 and 55.32 (2×OCH<sub>3</sub>); 83.38, 87.01, 87.65 and 99.38 (C=); 113.45 and 113.74 (2×C-iarom.); 114.03, 114.12, 134.16 and 134.51 (CH-arom.); 133.01 (C-5); 143.23 (CH-8); 142.43, 146.74 and 151.32 (C-2, C-4 and C-6); 160.48 and 160.99 (2×C-OMe). Exact mass (FAB HR MS) found: 423.1819; for C<sub>26</sub>H<sub>23</sub>N<sub>4</sub>O<sub>2</sub> [M+H] calculated: 423.1821.

3.2.8. 9-Isopropyl-2,6-bis[2-(4-methoxyphenyl)ethyl]purine (9). A solution of compound 8 (395 mg, 0.94 mmol) in dioxane (120 ml and EtOH (50 ml) was hydrogenated under atmospheric pressure in the presence of 5% Pd/C (200 mg) for 24 h at ambient temperature. The catalyst was filtered off on Celite and the solvents were evaporated. Column chromatography of the complex mixture gave compound 9 as yellow oil (55 mg, 14%). EI MS, m/z: 430 (37) [M], 387 (11), 277 (15), 149 (20), 121 (100). IR (CHCl<sub>3</sub>),  $\nu$ =1610, 1592, 1513, 1497, 1392, 1246 cm<sup>-1</sup>. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): 1.61 (d, 6H, J=6.8 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 3.13-3.16 (m, 4H, 2×CH<sub>2</sub>); 3.30-3.34 (m, 2H, CH<sub>2</sub>); 3.43-3.46 (m, 2H, CH<sub>2</sub>); 3.77 (s, 6H, 2×OCH<sub>3</sub>); 4.90 (sept., 1H, J=6.8 Hz, CH(CH<sub>3</sub>)<sub>2</sub>); 6.80 (d, 2H, J=8.6 Hz, H-arom.); 6.81 (d, 2H, J=8.5 Hz, H-arom.); 7.17 (d, 2H, J=8.5 Hz, H-arom.); 7.22 (d, 2H, J=8.5 Hz, H-arom.); 8.02 (s, 1H, H-8). <sup>13</sup>C NMR (100.6 MHz, CDCl<sub>3</sub>): 22.54 (CH(CH<sub>3</sub>)<sub>2</sub>); 33.50, 34.11, 35.17 and 41.08 (CH<sub>2</sub>); 46.90 (*C*H(CH<sub>3</sub>)<sub>2</sub>); 55.12 and 55.21 (2×OCH<sub>3</sub>); 113.66, 113.72, 129.40 and 129.43 (CH-arom.); 133.62, 133.95 and 136.82 (2×C-i-arom. and C-5); 140.86 (CH-8); 151.00, 157.79, 157.86, 161.17 and 164.08 (C-2, C-4 and C-6 and  $2 \times C$ -OMe). Exact mass (EI HR MS) found: 430.2358; for C<sub>26</sub>H<sub>30</sub>N<sub>4</sub>O<sub>2</sub> calculated: 430.2369.

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#### References

- (a) Veselý, J.; Havlíček, L.; Strnad, M.; Blow, J. J.; Donella-Deana, A.; Pinna, L.; Letham, D. S.; Kato, J.; Detivaud, L.; Leclerc, S. *Eur. J. Biochem.* **1994**, 224, 771–786. (b) Havlíček, L.; Hanuš, J.; Veselú, J.; Leclerc, S.; Meijer, L.; Shaw, G.; Strnad, M. *J. Med. Chem.* **1997**, 40, 408–412. (c) Legraverend, M.; Ludwig, O.; Bisagni, E.; Leclerc, S.; Meijer, L. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 793–798. (d) Legraverend, M.; Ludwig, O.; Bisagni, E.; Leclerc, S.; Meijer, L.; Giocanti, N.; Sadri, R.; Favaudon, V. *Bioorg. Med. Chem.* **1999**, *7*, 1281–1293. (e) Chang, Y.-T.; Gray, N. S.; Rosania, G.; Sutherlin, D. P.; Kwon, S.; Norman, T. C.; Sarohia, R.; Leost, M.; Meier, L.; Schultz, P. G. *Chem. Biol.* **1999**, *6*, 361–375.
- Verdugo, D. E.; Cancilla, M. T.; Ge, X.; Gray, N. S.; Chang, Y.-T.; Schultz, P. G.; Negishi, M.; Leary, J. A.; Bertozzi, C. R. *J. Med. Chem.* 2001, 44, 2683–2686.
- Chang, Y.-T.; Choi, G.; Bae, Y.-S.; Burdett, M.; Moon, H.-S.; Lee, J. W.; Gray, N. S.; Schultz, P. G.; Meier, L.; Chung, S.-K.; Choi, K. Y.; Suh, P.-G.; Ryu, S. H. *ChemBioChem* 2002, 879–901.
- Rosania, G. R.; Chang, Y.-T.; Perez, O.; Sutherlin, D.; Dong, H. L.; Lockhart, D. J.; Schultz, P. G. *Nat. Biotechnol.* 2000, *18*, 304–308.

- (a) Chang, Y.-T.; Wignall, S. M.; Rosania, G. R.; Gray, N. S.; Hanson, S. R.; Su, A. I.; Merlie, J.; Moon, H. S.; Sangankar, S. B.; Perez, O.; Heald, R.; Schultz, P. G. *J. Med. Chem.* 2001, 44, 4497–4500. (b) Perez, O. D.; Chang, Y. T.; Rosania, G.; Sutherlin, D.; Schultz, P. G. *Chem. Biol.* 2002, *9*, 475–483.
- Moon, H.-S.; Jacobson, E. M.; Khersonsky, S. M.; Luzung, M. R.; Walsh, D. P.; Xiong, W.; Lee, J. W.; Parikh, P. B.; Lam, J. C.; Kang, T.-W.; Rosania, G. R.; Schier, A. F.; Chang, Y.-T. *J. Am. Chem. Soc.* 2002, *124*, 11608–11609.
- Franěk, F.; Siglerová, V.; Havlíček, L.; Strnad, M.; Eckschlager, T.; Weigl, E. Collect. Czech. Chem. Commun. 2002, 67, 257–266.
- Hocek, M.; Holý, A.; Votruba, I.; Dvořáková, H. J. Med. Chem. 2000, 43, 1817–1825.
- Hocek, M.; Holý, A.; Votruba, I.; Dvořáková, H. Collect. Czech. Chem. Commun. 2001, 66, 483–499.
- Hocek, M.; Votruba, I. Bioorg. Med. Chem. Lett. 2002, 12, 1055–1058.
- For review on cross-coupling reactions in purines, see: Hocek, M. Eur. J. Org. Chem. 2003, 245–254.
- (a) Langli, G.; Gundersen, L. L.; Rise, F. *Tetrahedron* 1996, 52, 5625–5638. (b) Havelková, M.; Dvořák, D.; Hocek, M. *Synthesis* 2001, 1704–1710. (c) Hocek, M.; Holý, A.; Dvořáková, H. *Collect. Czech. Chem. Commun.* 2002, 67, 325–335.
- Otyepka, M.; Kryštof, V.; Havlíček, L.; Siglerová, V.; Strnad, M.; Koča, J. J. Med. Chem. 2000, 43, 2506–2513.
- Van Overmeire, I.; Boldin, S. A.; Venkataraman, K.; Zisling, R.; De Jonghe, S.; Van Calenbergh, S.; De Keukeleire, D.; Futerman, A. H.; Herdewijn, P. J. Med. Chem. 2000, 43, 4189–4199.